## Implementation, achievements and opportunities

## WP6

## A mutagenesis study aiming to establish C<sub>α</sub> hydroxylation activity at the active site of a pseudo enzyme

*Ba*3943 is a pseudo polysaccharide deacetylase (like YlxY and Ba0150) which displays no enzymatic and no hydroxylation activity. Basic properties of Ba3943:

- Hypothetical protein according to pubmed.com
- Belongs to the CE4 family and has a NodB domain according to Pfam
- Exhibits a 57% identity with YlxY from *B.subtilis*, they main conserved motifs of PDAs are only partially conserved in Ba3943.

		*		March 1997 August 2018		*				*
BA3943	113	VGLTINVAWG	166	VGNHSYTHPN	203	VRWFAPPSGS	231	WTVDTIDWK	259	IVLMHPTS
YlxY	132	VAFLINVAWG	185	IGNHSYNHPD	222	PKWFAPPSGS	250	WTVDTIDWQ	278	MILMHPTD
BA0150	59	VAFTFD I SWG	113	IGSMGYNYTS	150	IKLLRPPSGD	178	WSNNSNDWK	206	IVLLHASD
BC0361	200	IFVTFDDGMK	261	VQSHTATHAD	296	VIAI <mark>AYP</mark> FGH	324	TKPGQFIT.	338	LLKMKRVR
BA0424	70	IY <mark>LTFD</mark> NGYE	123	I GN H S W S H P D	161	VKYV <mark>RPP</mark> RGV	189	WSLAFLDWK	220	ILLHAIS
BsPdaA	68	IY <mark>LTFD</mark> NGYE	121	IGNHSFHHPD	159	NLYL <mark>RPP</mark> RGV	187	WSVAFVDWK	218	IY <mark>LLH</mark> TVS
BC1960	83	VALTEDDGPD	136	I <mark>g n h t</mark> y s <mark>h</mark> p n	173	PKFI <mark>RPP</mark> Y <mark>g</mark> e	201	WSVDTVDWK	229	VILQHSTP
BC1974	71	AYLTFDDGPG	123	V <mark>GMHSMTH</mark> NF	160	PKLT <mark>RPP</mark> YGS	191	WTIDSLDWR	226	VILMHDIH
SpPgdA	270	VALTEDDGPN	323	V <mark>gnhs</mark> ws <mark>h</mark> pi	360	SKLMRPPYGA	385	WDVDSLDWK	413	IV <mark>LMH</mark> DIH
ErPgd	285	IYLTFDDGPG	336	VAIHSASHKY	374	ASII <mark>RFP</mark> GGS	413	WNVSSGDAN	445	VVLQHDIK
SmPgdA	109	VFLTFDDGVD	163	L <mark>gihs</mark> fs <mark>h</mark> vy	207	TGVW <mark>RYP</mark> G <mark>G</mark> H	239	WNAAVGDAE	277	VVLMHDIS
10.000		Motif 1		Motif 2		Motif 3		Motif 4		Motif 5

Sequence alignment of the NodB domain (all 5 conserved motifs) of **BA3943** with YlxY, BA0150 (4M1B), BC0361 (4HD5), BA0424 (2J13), BsPdaA (1W17), BC1960 (4L1G), BC1974 (5N1J), SpPgdA (2C1G), ErPgd (5JMU) and SmPgdA (2W3Z). The PDB id codes are in parentheses. Inactive PDAs are depicted in black, BC0361 (unknown substrate, PG binding) in orange, MurNAc PDAs in red and GlcNAc PDAs in blue.

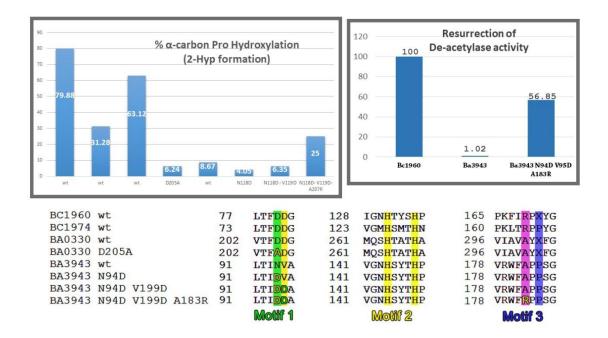
- BA3943 and BA0150 structurally resemble to MurNAc deacetylases
- <u>Similarities</u>: BA3943 retains the catalytic histidine (MT5), the metal-binding histidines (MT2) and the conserved proline (MT3)
- <u>Differences</u>: BA3943 does not retain the catalytic aspartic acid which is replaced by asparagine (MT1), the metal-binding aspartic acid which is replaced by valine (MT1) and the, very significant for both deacetylation and hydroxylation activities, arginine which replaced by alanine (MT3)
- Inactive against glycol-chitin, chitooligomers and PG isolated from *E.coli*

On the basis of sequence-structure-activity relationships established in the project, a number of mutants were engineered and tested. The aim was to restore the essential motifs associated with enzymatic and hydroxylation activity. The following mutants of Ba3943 were engineerd as steps towards restoration of sequence motifs and establishment of hydroxylation/ catalytic activity: Ba3943 N94D, Ba3943 N94D V95D, Ba3943 N94D V95D A183R. Several other mutants were also produced to examine specific structural or hydroxylation aspects of the Ba3943 pseudoenzyme.

ba3943 and the mutant genes, were expressed in E.coli and the recombinant protein was purified to near homogeneity. Heavy aggregation was a common property of

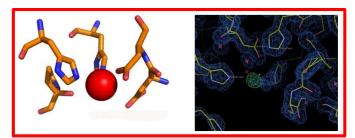
purified proteins. Hydroxylation activity was examined through the detection of 2-Hyp by MS or by X-ray crystallography. Enzymatic activity was examined through incubation with commonly used deacetylase substrates. BA3943, Ba3943 N94D and Ba3943 N94D V95D were enzymatically inactive against radio labeled glycol chitin, chito-oligomers and peptidoglycan isolated from *E.coli*, when tested in a wide range of pH and in the presence or absence of the divalent cations  $Co^{+2}$ ,  $Zn^{+2}$ ,  $Ni^{+2}$  and  $Mn^{+2}$ . A 2-Hyp residue could not be detected either. On the other hand, the triple mutant Ba3943 N94D V95D A183R exhibits a significant enzymatic and hydroxylation activity.

Ba3943 (wt protein) inactive
Ba3943 N94D inactive
Ba3943 N94D V95D inactive
Ba3943 N94D V95D A183R active



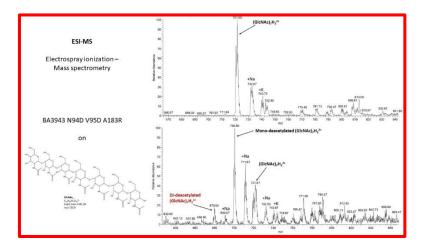
The catalytic activity of Ba3943 N94D V95D A183R in the presence of metals exhibits a maximum value for  $Co^{+2}$ , a common property of PDAs, despite the fact that the natural catalytic metal for these enzymes is  $Zn^{+2}$ .

With the exception of Ba3943 N94D V95D A183R, no metal could be detected from electron density maps in Ba3943 and its mutants. However, even for the activated mutant Ba3943 N94D V95D A183R, the  $Zn^{+2}$  occupation of the metal binding site is low.

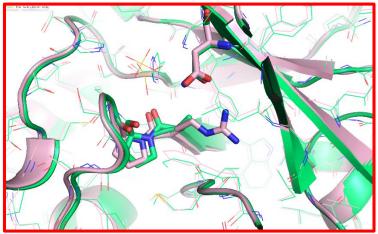


Right: Electron density maps of Ba3943 N94D V95D A183R calculated in the region corresponding to the catalytic/ hydroxylation site of PDAs (blue:  $2F_o$ - $F_c$ , green:  $F_o$ - $F_c$  maps, contoured at  $2\sigma$  and  $4\sigma$  respectively). Left: The probable  $Zn^{+2}$  (red) binding site of Ba3943 N94D V95D A183R, modelled according to the electron density.

Catalytic activity of the Ba3943 N94D V95D A183R mutant was detected using electrospray ionization-mass spectrometry:

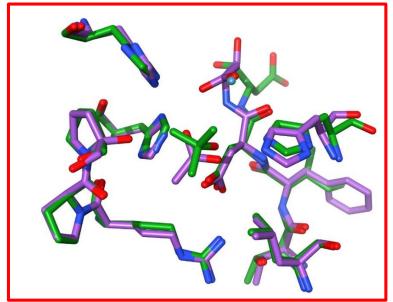


All structural differences between Ba3943 and Ba3943 N94D V95D A183R, cluster in the active/hydroxylation site of the mutant, as shown in the following figure:



Superposition of the active/ hydroxylation site of Ba3943 N94D V95D A183R with that of Ba3943 (green)

The crystal structure of the "resurrected" Ba3943 N94D V95D A183R, reveals a striking structural similarity in the potential hydroxylation/ catalytic site with corresponding site of fully active PDAs, e.g. Bc1960:



Superposition of the active/ hydroxylation site of Bc1960 with the engineered site of Ba3943 N94D V95D A183R.